## Genomic Psychiatry

### **THOUGHT LEADERS INVITED REVIEW**

# **Genomic Psychiatry**

# Roles of alternative polyadenylation in psychiatric disorder risk

Michelle Paff<sup>1</sup> <sup>®</sup>[,](https://orcid.org/0000-0001-9931-5990) Steven F. Grieco<sup>2,[3](#page-0-2)</sup> ®, Ya Cui<sup>3</sup> ®, Ole A. Andreassen<sup>4</sup> ®, Albert LaSpada<sup>[6,](#page-0-4)[7,](#page-0-5)8</sup>, Wei Li, PhD<sup>6</sup> ®, and Xiangmin Xu<sup>2,3,[8,](#page-0-6)[9,](#page-0-7)[10,](#page-0-8)[11](#page-0-9)</sup>

**Alternative polyadenylation (APA) is a pervasive regulatory mechanism in the human brain that controls the stability and cellular localization of mRNA transcripts. Single-nucleotide polymorphisms associated with psychiatric disorders may exert their deleterious effects by altering 3' untranslated site usage, which may change the stability and processing of mRNA transcripts. The authors previously performed a 3'APA transcriptomic-wide association study using the DePars2 framework and the GTEx v8, PsychENCODE, and ROS/MAP datasets to identify APA-linked genes associated with eleven brain disorders. Here we focus on 3'APA-linked genes associated with the major psychiatric conditions: schizophrenia, bipolar disorder, and depression. There are 286 APA-linked genes associated with these psychiatric disorders, and 60%–65% of these genes have not been associated with the major psychiatric disorders through their expression and/or splicing. Protein–protein interaction networks indicate that APA-linked genes associated with schizophrenia are involved in intracellular transport and cellular localization pathways. Future research is needed to elucidate the role of alternative 3' untranslated region usage of APA-linked genes on neuronal function and phenotypic expression in psychiatric disorders.**

*Genomic Psychiatry* (2024) **1**, 1–8; doi: [https://doi.org/10.61373/gp024i.0049;](https://doi.org/10.61373/gp024i.0049) Published online: 18 July 2024. **Keywords:** Alternative polyadenylation, transcriptome-wide association studies, psychiatric disorders, schizophrenia, bipolar disorder, depression

#### **Introduction**

Over the past 15 years, genome-wide association studies (GWAS) have uncovered hundreds of single-nucleotide polymorphisms (SNPs) associated with the major psychiatric disorders. However, translating these genetic associations to biologically relevant mechanisms remains a major challenge. The majority of GWAS-significant SNPs localize to non-coding regions of the genome and often reside within loci demonstrating a high degree of linkage disequilibrium. Thus, identifying causal genes responsible for the phenotypic expression of psychiatric conditions can be an arduous process. Transcriptome-wide association studies (TWAS) serve as one method of nominating putative causal genes by integrating populationlevel transcriptomic datasets and GWAS summary statistics [\(1–](#page-5-0)[4\)](#page-5-1). Leveraging population-level datasets with both genotype and gene expression information, TWAS impute the *cis*-component of gene expression of common variants to prioritize genes at trait-associated loci, referred to as quantitative trait loci (QTL).

While the majority of TWAS have focused on total gene expression, it has become increasingly realized that disease-relevant SNPs may have regulatory effects that alter transcript splicing, epigenetic regulation, and protein expression, among many other potential mechanisms  $(5, 6)$  $(5, 6)$  $(5, 6)$ . Recently, alternative 3' untranslated polyadenylation site usage has been recognized as a mechanism by which GWAS significant SNPs may affect the abundance of transcript isoforms  $(7, 8)$  $(7, 8)$  $(7, 8)$ . Alternative polyadenylation (APA) is a pervasive regulatory mechanism of mRNA trafficking and translation that is particularly critical in the central nervous system (CNS). Aberrant APA plays a role in multiple neurological disorders, including Parkinson's disease (PD)  $(9)$ , Huntington's disease  $(10)$ , and certain forms of intellectual disability [\(11\)](#page-5-8). Unlike total expression levels or alternatively spliced mRNA isoforms, the impact of APA on disease-related

gene expression has not been accounted for in the majority of TWAS analyses.

Polyadenylation is an important step in the production of mature mRNA species. Following transcription, the 3' end of a mRNA is cleaved at either a proximal or distal 3' APA site, and a poly(A) tail is synthesized at the 3' terminus. APA sites are located within the 3' untranslated regions (3'UTRs) of genes. Approximately 70% of human genes include multiple APA sites that produce 3'UTRs of different lengths [\(12\)](#page-5-9). Use of a proximal APA site produces a shorter mRNA with a short 3'UTR, while use of a distal APA site produces a longer mRNA with a long 3'UTR [\(Figure 1A\)](#page-1-0). 3'UTRs contribute to posttranscriptional regulation of gene expression in multiple ways. Regulatory elements residing within 3'UTRs affect mRNA stability, translation efficiency, and cellular localization [\(13\)](#page-5-10). 3'UTRs produced by distal APA sites tend to contain more target sequences for RNAbinding proteins and microRNAs (miRNA), which can destabilize mRNAs and promote degradation [\(Figure 1B\)](#page-1-0) [\(14,](#page-5-11) [15\)](#page-5-12). As longer 3'UTRs tend to contain a greater number of regulatory elements, genetic mutations that lead to a change in APA site usage can alter mRNA stability.

<span id="page-0-6"></span><span id="page-0-5"></span><span id="page-0-2"></span>APA may serve as a regulatory mechanism with important developmental implications. As cells and tissues evolve from primitive to fully differentiated forms, the APA of their expressed transcripts often changes. Transcriptomic studies of embryonic mouse tissue have shown that mouse genes tend to express transcripts with longer 3'UTRs as embryonic development progresses [\(16\)](#page-5-13). Additionally, 3'UTR length varies according to cell type. Amongst the major cell types, stromal cells and neuronal cell types express transcripts with the longest 3'UTRs, while blood cells, hepatocytes, chondrocytes, and osteoblasts express transcripts with the short-est 3'UTRs [\(17\)](#page-5-14). Furthermore, among cells of a given lineage, use of long 3'UTRs tends to increase as primitive cells differentiate to mature cell

<span id="page-0-9"></span><span id="page-0-7"></span>**Corresponding Author:** Michelle Paff, MD; University of California, Irvine, Department of Neurological Surgery, 200 South Manchester Avenue, Suite 210, Orange, CA 92868, Phone: 714-456-6966, Fax: 714-456-6966. E-mail: [mpaff@hs.uci.edu](mailto:mpaff@hs.uci.edu)

Received: 15 April 2024. Revised: 28 May 2024. Accepted: 19 June 2024.

<span id="page-0-8"></span><span id="page-0-4"></span><span id="page-0-1"></span>

<span id="page-0-3"></span><span id="page-0-0"></span><sup>&</sup>lt;sup>1</sup>Department of Neurosurgery, School of Medicine, University of California, Irvine Medical Center, Orange, California 92868, USA; <sup>2</sup>Department of Anatomy and Neurobiology, School of Medicine, University of California, Irvine, California 92697, USA; <sup>3</sup>Center for Neural Circuit Mapping, University of California, Irvine, California 92697, USA; <sup>4</sup>Division of Computational Biomedicine, Department of Biological Chemistry, School of Medicine, University of California, Irvine, Invine, California 92697, USA; <sup>5</sup>Norwegian Centre for Mental Disorders Research (NORMENT), Division of Mental Health and Addiction, Oslo University Hospital, Oslo 0450, Norway; <sup>6</sup>Department of Biological Chemistry, School of Medicine, University of California, Irvine, California 92697, USA; <sup>7</sup>Departments of Pathology and Laboratory Medicine, Neurology, and Biological Chemistry, University of California Irvine, Irvine, California 92697, USA; <sup>8</sup>UCI Institute for Neurotherapeutics, University of California Irvine, Irvine, California 92697, USA; <sup>9</sup>Department of Microbiology and Molecular Genetics, University of California, Irvine, California 92697, USA; <sup>10</sup>Department of Biomedical Engineering, University of California, Irvine, California 92697, USA; <sup>11</sup>Department of Computer Science, University of California, Irvine, California 92697, USA

<span id="page-1-0"></span>



**Figure 1.** Alternative polyadenylation (APA). (**A**) Splicing can lead to mRNA isoforms with different 3' UTRs and/or APA based on is polyadenylation site (PAS) usage in various combinations. (B) Two mRNA isoforms with different 3' UTRs due to APA. Only the constitutive UTR (cUTR) is present in the short isoform, whereas the the alternative UTR (aUTR) is also present in the long isoform. Interactions between the UTRs and RNA-binding proteins (RBPs), miRNAs and long non-coding RNAs (IncRNAs) can have functional consequences. **(C**) BDNF 3' UTR-APA isoform localization in neurons supports dendritic protein synthesis—the long isoform localizes to dendrites more than the short isoform (adapted from Tian and Manley, 2017, Nat. Rev. Mol. Cell Biol).

types. For example, in the hematopoietic lineage, the switch from short to long 3'UTRs is explained by the evolution from primitive to definitive erythropoiesis  $(17)$ . Thus, APA is a dynamic process that is particularly critical during prenatal development.

Changes in APA site usage may alter the distribution and localization of translated proteins. For example, the short isoform of brain-derived neurotropic factor (BDNF) is restricted to the neural cell body while the long isoform localizes to dendrites [\(Figure 1C\)](#page-1-0) [\(18\)](#page-5-15). The role of 3'UTRs in influencing the cellular localization of mRNA is particularly consequential for neurons since dendrites, axon terminals and cell bodies have distinct roles in health and disease. APA of the alpha synuclein (aSyn) transcript was implicated in a familial form of PD [\(9\)](#page-5-6). For example, SNPs associated with a familial form of PD were found to increase the abundance of the long 3'UTR isoform of alpha synuclein relative to the short 3'UTR isoform. Overproduction of the long 3'UTR isoform was found to lead to the accumulation of alpha synuclein protein with the mitochondria of neurons, leading to neuronal dysfunction. Thus, genetic variants that alter APA site usage may have a significant impact on cellular homeostasis, particularly in the brain where the majority of transcripts are regulated by APA.

#### **APA in Psychiatric Conditions**

The CNS harbors the longest 3'UTRs of any tissue, suggesting that regulatory elements within 3'UTRs play an important role in protein expression throughout the brain. While traditional TWAS methods have identified mRNA expression levels and isoform level changes due to alternative splicing, they have not investigated APA. To address this gap, Cui *et al*. [\(8\)](#page-5-5) performed a 3'UTR TWAS to identify local genetic effects associated with variation of 3'UTR usage among the GTEx  $v8$  [\(19\)](#page-5-16), ROS/MAP [\(20\)](#page-5-17), and psychENCODE [\(21\)](#page-5-18) datasets. The authors implemented the DaPars2 framework  $(22)$  to calculate the percentage of distal poly $(A)$  site usage and to identify 3'UTR lengthening and shortening events. The DePars2 framework calculates a 3'UTR usage value for each transcript across samples. A linear regression framework is then applied to test the association between normalized values of 3'UTR usage and SNPs within an interval of 1Mbp of the 3'UTR region, adjusting for covariates. They identified *cis*-SNPs associated with 3'UTR usage and examined the association between GWAS summary statistics and 3'UTR usage. Transcriptome and individual-matched genotype data from the ROS/MAP, PsychENCODE, and GTEx Consortia was then implemented to establish 3'aTWAS single-tissue prediction models for 3'UTR usage using FUSION [\(2\)](#page-5-20). They discovered 354 APA-linked disease susceptibility genes identified among 11 brain disorders, including the major psychiatric disorders, schizophrenia (SCZ), bipolar disorder (BD), and depression (DEP) [\(Figure 2A–C\)](#page-2-0). The largest number of 3'aTWAS-signficant genes was found for SCZ, with 281 non-HLA APA-linked genes associated with the disorder among the three datasets. A comprehensive list of APA-linked genes for SCZ, BD, and DEP from Cui et al. can be found in Supplementary Table S1.

Some of the genetic risk loci associated with APA (3'aQTLs) were also implicated in expression and splicing TWAS. Many 3'aTWAS genes had a more significant 3'aQTL signal than eQTL or sQTL signals, indicating that their GWAS signal is better explained by their effect on APA. One validated example of this is the detection of a known APA-susceptibility gene, SNCA (encoding aSyn) associated with PD. SNCA was the most significant gene identified in all three reference panels, and longer 3'UTR usage was associated with increased PD risk, consistent with prior evidence [\(9\)](#page-5-6). The leading PD GWAS SNP near *SNCA* is less strongly associated with differential expression and splicing, supporting that 3'UTR usage is primarily responsible for PD risk. Among the major psychiatric disorders, 49 of 151 (32%) non-HLA, APA-linked genes associated with SCZ were also implicated in expression TWAS, splicing TWAS, or both. Eighty-seven genes, including *ZFN592*, *PBX2* and *RBX1*, were associated with SCZ only through APA and not previously implicated in other TWAS [\(Figure 3A\)](#page-2-1). Seventeen of 71 (24%) APA-linked genes associated with BD were also implicated in expression and splicing TWAS [\(Figure 3B\)](#page-2-1), and 24 of 64 (37.5%) APAlinked genes associated with DEP were also implicated through expression and/or splicing [\(Figure 3C\)](#page-2-1).

In the following paragraphs, we will discuss the implications of some APA-linked genes, identified in Cui *et al*. associated with the major psychiatric conditions, SCZ, BD, and DEP.

#### *DDHD2* **(DDHD-Domain-Containing 2)**

The *DDHD2* gene on chromosome 8p encodes a triacylglyceride hydrolase that is involved in membrane trafficking between the endoplasmic reticulum and Golgi body. *DDHD2* is ubiquitously expressed in the brain, and multiple transcript variants result from alternative splicing. Missense mutations in *DDHD2* cause an autosomal recessive form of hereditary spastic paraplegia, which includes intellectual disability among its clinical features [\(23\)](#page-6-0). Furthermore, *DDHD2* knockout mice demonstrate motor and cognitive deficits as well as lipid accumulation within neurons [\(24\)](#page-6-1). Although the role of *DDHD2* in cognitive function has not been fully elucidated, *DDHD2* has been linked to caudate, putamen, and pallidum volume and is downregulated in the dorsolateral prefrontal cortex (DLPFC) of patients with SCZ [\(25\)](#page-6-2). The 8p12 genomic region, near the *DDHD2* gene, has been identified as a significant risk locus for SCZ among Han Chinese and European populations  $(26, 27)$  $(26, 27)$  $(26, 27)$  as well as for autism spectrum disorder [\(28\)](#page-6-5) and BD [\(6,](#page-5-3) [29\)](#page-6-6). Decreased expression of *DDHD2* has been associated with SCZ in multiple TWAS [\(6,](#page-5-3) [30\)](#page-6-7), including eQTLs derived from prenatal brain and dopaminergic neurons [\(6,](#page-5-3) [8,](#page-5-5) [29,](#page-6-6) [31\)](#page-6-8).

#### [gp.genomicpress.com](https://gp.genomicpress.com)

<span id="page-2-0"></span>

**Figure 2.** Psychiatric 3'aTWAS. (**A1**) RNA sequencing and matched genotype data were collected from the GTEx, ROS/MAP, and PsychENCODE cohorts as reference panels. (A2) 3'aQTL analysis was performed, and then a 3'aTWAS model (A3) was built to predict the APA usage of target genes with cis-SNPs in the reference panels. (**B1**) We used GWAS summary statistics and the (**B2**) 3'aTWAS models for each reference panel to (**B3**) perform 3'aTWAS analysis to nominate susceptibility genes in brain disorders. **(C1**) APA-linked susceptibility genes in brain disorders identified by 3'aTWAS (only PsychENOCDE data shown). **(C2**) Bar plot shows the number of 3'aTWAS significant genes for 11 brain disorders in PsychENCODE DLPFC. **(C3**) Venn diagram shows the overlap of 3'aTWAS significant genes for 11 brain disorders with expression and splicing TWAS (includes the GTEx, ROS/MAP, and PsychENCODE cohorts) (adapted from Cui *et al*., 2023, Nat Commun).

<span id="page-2-1"></span>





In the 3'aTWAS analysis, the short 3'UTR isoform of *DDHD2* was significantly associated with SCZ in the GTEx v8, ROS/MAP, and psychEN-CODE datasets. This is consistent with previous analyses which identified a GWAS-significant SNP within the 3'UTR of the *DDHD2* mRNA transcript that disrupts binding of the quaking RNA-binding protein [\(32\)](#page-6-9). Interestingly, downregulation of the quaking RNA-binding protein was associated with risk of SCZ in a large Swedish pedigree, indicating that the RNAbinding protein target sequence within the 3'UTR of *DDHD2* plays a role in risk of SCZ [\(33\)](#page-6-10). Elimination of target sequences for RNA-binding proteins, such as that for the quaking RNA-binding protein, may occur with use of proximal 3'UTR sites that omit portions of the extended 3'UTR, yielding a similar result.

#### *ARL3* **(ADP Ribosylation Factor-Like GPTase 3)**

The product of the *ARL3* gene on chromosome 10q is a GTP-binding protein that localizes to cilia and microtubules and plays a role in the formation of axons and cilia. Mutations in *ARL3* can result in ciliopathies, including neurodevelopmental disorders such as Joubert syndrome, characterized by hypoplasia of the cerebellar vermis, brainstem abnormalities, psychomotor delay, hypotonia, and retinal abnormalities [\(34\)](#page-6-11). A SNP located within an intron of *ARL3* was significantly associated with SCZ in a Han Chinese population [\(35\)](#page-6-12), and decreased expression of *ALR3* has been associated with SCZ in TWAS  $(6, 31)$  $(6, 31)$  $(6, 31)$  and in a proteome-wide association study [\(36\)](#page-6-13). The short 3'UTR isoform was associated with SCZ in all three datasets examined in Cui *et al*., however, the biological significance of increased short 3'UTR usage by the *ARL3* gene in regards to either ciliary function or neuronal development has yet to be explored.

#### *SNX19* **(Sorting Nexin 19)**

*SNX19* on chromosome 11q encodes a protein that belongs to a family of sorting nexins that function in endosomal trafficking regulation and sorting. Expression and alternative splicing of *SNX19* has been associated with SCZ in multiple TWAS [\(6,](#page-5-3) [37](#page-6-14)[–39\)](#page-6-15), while short 3'UTR usage of *SNX19* was associated with SCZ in transcriptomic data from multiple brain regions of the GTEx v8 dataset. Greater expression of an isoform with skipping of exon 9 was associated with a downstream SCZ risk locus, with most unaffected individuals expressing very low levels of this transcript isoform. Skipping of exon 9 produces a frameshift that is predicted to result in the absence of the sorting C-terminal domain  $(38)$ . In situ hybridization studies support that *SNX19* is localized to glutamatergic neurons in the DLPFC [\(40\)](#page-6-17). *SNX19* may be associated with the sodium-coupled neutral amino acid transporter 1 (SLC38A1), which supplies neurons with glutamine for synthesis of neurotransmitters [\(40\)](#page-6-17). The impact of short 3'UTR usage of *SNX19* on neuronal function has yet to be investigated.

#### *ZNF592* **(Zinc Finger RNA-binding Protein 2)**

The *ZNF592* gene encodes a 1,267 amino acid zinc finger protein that is expressed in the CNS [\(41\)](#page-6-18). Zinc finger proteins function as transcriptional regulators, mediating interactions between DNA and proteins. Missense mutations in *ZNF592* cause cerebellar ataxia with mental retardation, optic atrophy, and skin abnormalities [\(41\)](#page-6-18). In the 3'aTWAS, short 3'UTR usage of *ZNF592* was associated with both SCZ and BD in the ROS/MAP and psychENCODE datasets. Importantly, *ZNF592* has not been previously identified in expression or splicing TWAS, indicating that the GWAS signal at this locus is almost entirely explained by alternative polyadenylation.

#### *FADS1* **(Fatty Acid Desaturase 1)**

*FADS1* encodes a fatty acid desaturase, which is a rate limiting enzyme in desaturation of long-chain polyunsaturated fatty acids. The *FADS1* and *FADS2* genes in 11q12.2 locus are in tight LD, and this locus has been associated with BD in multiple populations. A GWAS significant SNP at the 11q12.2 locus, containing the *FADS1/2* gene, was associated with BP in Asian populations [\(42,](#page-6-19) [43\)](#page-6-20) and then replicated in GWAS of European populations.[\(44,](#page-6-21) [45\)](#page-6-22) In Cui *et al*., expression, alternative splicing, and short 3'UTR usage of *FADS1* was associated with BP in the ROS/MAP dataset. Multiple mRNA isoforms of *FADS1* are generated by alternative transcript initiation, alternative polyadenylation site usage, and internal exon deletions [\(46\)](#page-6-23). There are seven possible isoforms of *FADS1* that differ only in the length of their poly(A) tail due to use of alternative poly(A) sites.

Since the commencement of crop agriculture leading to increased intake of grain oils, there has been an increase in the proportion of individuals carrying a haplotype associated with greater *FADS1/2* activity that has a protective effect against BD [\(47\)](#page-6-24). Transgenic mice with decreased *FADS1/2* activity demonstrate behavioral changes including bouts of hyperactivity interspersed with periods of depressive-like hypoactivity as well as abnormal circadian rhythms, suggesting that reduced FADS1/2 activity produces a robust animal model of BD  $(48)$ . Furthermore, these behavioral deficits were rescued with dietary supplementation of polyun-saturated fatty acids [\(48\)](#page-6-25). However, dietary supplementation with omega-3 fatty acids to treat mood fluctuations in BD have demonstrated mixed results [\(49,](#page-6-26) [50\)](#page-6-27).

#### *GABRA2* **(Gamma-Aminobutyric Acid Type A Receptor Alpha 2)**

*GABRA2* encodes the alpha subunit of the GABAA receptor, which mediates anxiolytic-like, reward-enhancing, and anti-hyperalgesic actions of benzodiazepines [\(51\)](#page-6-28). *GABRA2* has been implicated in alcoholism and substance abuse disorders [\(52,](#page-6-29) [53\)](#page-6-30), and SNPs residing within GABAA receptor subunit genes have been associated with risk of BD [\(54](#page-6-31)[–56\)](#page-6-32). GABA receptor mRNAs display long 3'UTRs, and increased 3'UTR length is linked to reduced translation [\(57,](#page-6-33) [58\)](#page-6-34). In our 3'aTWAS, short 3'UTR usage was associated with BD in the GTEx v8, ROS/MAP, and psychENCODE datasets. Strong 3'aQTLs but weak eQTLs were found for *GABRA2* in 3'aTWAS, indicating that the 3'aQTL association with *GABRA2* almost entirely explains the BD GWAS signal at this locus.

#### *CACAN1B* **(Calcium Voltage-gated Channel Subunit Alpha 1B)**

Genes encoding subunits of calcium channels have been repeatedly implicated in BD GWAS [\(44,](#page-6-21) [59,](#page-6-35) [60\)](#page-6-36). *CACNA1B* encodes the alpha-1B subunit; the pore-forming subunit, of the presynaptic N-type voltagedependent calcium channel (Ca<sub>V</sub>2.2). Two mRNA isoforms of *CACNA1B* are produced by alternative splicing based on the inclusion of one of two mutually exclusive exons, e37a and e37b, which encode se-quences of the C-terminus [\(61\)](#page-6-37). Compared to channels containing e37b, channels containing e37a are trafficked more efficiently to the cell membrane and are inhibited more strongly by G-protein–coupled receptors [\(62,](#page-7-0) [63\)](#page-7-1). E37a-rich channels are abundant in calcium/calmodulindependent excitatory projection neurons, including those comprising excitatory cortico-hippocampal synapses [\(64\)](#page-7-2). Mice expressing only the e37b isoform display decreased novelty-induced anxiety, suggesting that alternative splicing of *CACN1B* influences withdrawal and anxiety-related behaviors.

It has been demonstrated in sympathetic neurons that the half-life of the alpha-1B subunit mRNA is regulated by its 3'UTR and modulated by voltage-dependent calcium entry [\(65\)](#page-7-3). In the analysis by Cui *et al*., short 3'UTR usage, expression, and splicing of *CACNA1B* were all associated with BD in the GTEx v8 and ROS/MAP cohorts.

The replicated association of multiple calcium channel subunit genes with BD has prompted the investigation of calcium channel blockers as adjunctive therapy in the treatment of BD, however, with about half of patients demonstrating a clinical benefit [\(66\)](#page-7-4). In a small cohort of 38 patients with BD, two SNPs in the *CACNA1B* locus were associated with treatment response to calcium channel blockers. However, no significant difference in the predicted expression of *CACNA1B* was found between responders and non-responders [\(67\)](#page-7-5).

#### *ARL17A* **(ADP Ribosylation Factor Like GTPase 17A)**

*ARL17A* encodes a GTP-binding protein which is a member of the ADPribosylation factor family. ADP ribosylation factor like GTPase 17A plays a role in vesicle-mediated intracellular protein transport between the endoplasmic reticulum and the Golgi apparatus and is important for neuronal development. Long 3'UTR usage of ARL17A was significantly associated with both SCZ and DEP among the GTEx v8, ROS/MAP, and psychENCODE datasets. Although the contribution of *ARL17A* to psychiatric conditions has yet to be fully elucidated, expression of *ARL17A* in the DLPFC, putamen, and cerebellum has been implicated in SCZ [\(68\)](#page-7-6). *ARL17A* expression has additionally been associated with intracranial brain volume [\(69\)](#page-7-7), thalamic volume in childhood [\(68\)](#page-7-6), and reaction time and cognitive function [\(69\)](#page-7-7).



<span id="page-4-0"></span>



**Figure 4.** Protein–protein interaction networks for 3'aTWAS significant genes for SCZ (**A**), BD (**B**), and DEP (**C**). Pathway analysis demonstrates that 3'aTWAS significant genes associated with SCZ are enriched in intracellular transport and cellular localization pathways.

#### *MTCH2* **(Mitochondrial Carrier Homolog 2)**

*MTCH2* encodes a member of the SLC25 family of nuclear-encoded transporters, which is localized to the outer mitochondrial membrane and plays an important role in oxidative phosphorylation [\(70\)](#page-7-8). *MTCH2* has been associated with body-mass index in multiple obesity GWAS [\(71,](#page-7-9) [72\)](#page-7-10) as well as with neuroticism [\(73\)](#page-7-11) and susceptibility to loneliness [\(74\)](#page-7-12). The SNP implicated in neuroticism was found to regulate *MTCH2* in the cerebellum [\(73\)](#page-7-11). Long 3'UTR usage by *MTCH2* was significantly associated with DEP in all three reference panels. Manjunath and colleagues [\(75\)](#page-7-13) demonstrated that transcription of *MTCH2* is subject to stop codon readthrough in which three different transcript isoforms may be produced depending on which stop codon is used. The long isoform localizes to the cytoplasm where it is rapidly degraded, and this leads to reduced mitochondrial membrane potential and decreased production of reactive oxygen species [\(75\)](#page-7-13). While APA of the *MTCH2* has not been studied, APA may similarly regulate the cellular localization of the *MTCH2* transcript and consequently alter mitochondrial function in neurons.

#### **Protein Interaction Networks**

We used STRING [\(76\)](#page-7-14) to evaluate the interconnectivity of non-HLA 3'aT-WAS genes by physical protein–protein interactions (PPIs) and Cytoscape (v3.3.0) [\(77\)](#page-7-15) to visualize the PPI networks for APA-linked genes associated with SCZ, BD, and DEP [\(Figure 4\)](#page-4-0). APA-linked genes associated with the major psychiatric conditions are enriched in biological pathways, including intracellular transport ( $p = 0.0152$ ) and establishment of localization in a cell ( $p = 0.0152$ ). Nodes in the SCZ PPI network, the

largest network, include genes involved in pre-mRNA splicing, such as *SNU1*, *SMU1*, *TXNL4A*, and *ZMAT2*. Another prominent node is centered on *RBX1*, an E3 ubiquitin protein ligase, which interacts with *GLMN*, an ubiquitin ligase inhibitor, and *CUL3*, a component of ubiquitin protein ligase complexes. For the DEP PPI network, the most prominent node is centered on *MAPT*, microtubule-associated tau protein, which functions in promoting microtubule assembly and stability. While 3'aTWAS significant genes represent a small portion of SCZ QTLs, pathways are notably distinct from those that have been associated with differentially expressed and spliced genes, such as metabolic pathways, synaptic plasticity, excitatory synapses, and immune-related pathways [\(5,](#page-5-2) [78\)](#page-7-16). A summary of the prominent protein–protein interaction networks identified in this analysis is provided in Supplementary Table S2.

#### **Conclusions and Future Directions**

In the post-GWAS era, the field of psychiatric genomics is challenged with interpreting the biological significance of hundreds of risk loci. TWAS has become a widely implemented method of nominating putative causal genes by leveraging relatively small transcriptomic datasets. However, there are multiple mechanisms by which SNPs may contribute to a biological phenotype, which are not all considered in most TWAS analyses. The discovery of 3'aQTLs helps explain some GWAS-significant SNPs that are not associated with differential expression or splicing in traditional TWAS analyses. While a percentage of APA-linked genes are also implicated in expression and splicing TWAS analyses, approximately 60%–75% of APAlinked genes for SCZ, BD, DEP have not been associated with total gene

**Thought Leaders Invited Review Paff et al.**



expression or isoform expression in the largest TWAS. For example, GWASsignificant SNPs, rs2024566 and rs5751204, on chromosome 22 near the *SNU13* gene did not reach significance in a large isoform-level TWAS [\(6\)](#page-5-3); however, alternative 3'UTR usage of this gene was associated with SCZ in the analysis of Cui *et al*., which utilized the same transcriptomic datasets. The association of genes with differential 3'UTR usage in 3'aTWAS suggests that aberrancies in mRNA translocation or degradation may play a role in disease risk. However, further confirmatory studies are needed to determine the impact of differential 3'UTR usage of 3'aTWAS significant genes on neuronal function and homeostasis. As transcriptomic reference sets continue to grow, it may eventually become possible to directly measure differential expression, splicing, 3'UTR usage between cases and controls. This will be critical for validating the findings of TWAS and guiding focused efforts to decipher downstream mechanisms.

In addition to APA of SNCA in PD, there several well-studied examples of APA alterations having significant neurological consequences that help explain phenotypic traits. A relatively well-characterized example is the MECP2 gene, which encodes methyl-CpG binding protein 2 (MeCP2), involved in the regulation of transcription of many different genes. Loss-of-function mutations of MECP2 result in Rett syndrome, which is characterized by developmental regression and intellectual disability beginning at around 18 months of age [\(79\)](#page-7-17). The MECP2 transcript exists in two isoforms with either a long 3'UTR or short 3'UTR [\(80\)](#page-7-18). The proximal APA site of the gene, which produces the short isoform, is increasingly used throughout postnatal development and is associated with increased protein abundance  $(81)$ . The long MECP2 isoform, on the other hand, is translated less efficiently compared to the short isoform and leads to decreased protein abundance. Mutations in gene products that regulate APA of MECP2 can also cause forms intellectual disability and autism spectrum disorders. An example of this is copy number variants (CNV) duplications of NUDT21 [\(11\)](#page-5-8). The protein product of NUDT21, a component of mammalian cleavage factor 1 complex (CFIm25), binds to the distal APA sites of genes, including MECP2, and facilitates production of transcripts with long 3'UTRs [\(82\)](#page-7-20). Lymphoblastoid cell lines derived from patients with NUDT21 duplications have ∼50% less MeCP2 protein and increased abundance of the long MECP2 transcript [\(82\)](#page-7-20). In this scenario, the reduction in MeCP2 protein is not quite as severe as in Rett syndrome, yet it is sufficient to produce intellectual disability [\(11\)](#page-5-8).

More detailed investigations as described above are needed to examine the impact of APA of genes associated with psychiatric disorders. As we learn more about how APA alterations affect the brain, we will better define the molecular and biological underpinnings of neuropsychiatric disorders, which will guide the development of treatment strategies. Efforts to determine the critical stages of development during which APA alterations contribute to phenotypic expression will also be informative for identifying opportunities for therapeutic intervention.

#### **Author Contributions**

M.P. performed the literature review and wrote the manuscript. S.G. conceptualized the review, edited the manuscript and produced the figures for the manuscript. Y.C. contributed content for the review and edited the manuscript. O.A.A. reviewed and edited the manuscript. A.L.S. supervised preparation and edited the manuscript. W.L. conceptualized and edited the manuscript. X.X. supervised and edited the manuscript.

#### **References**

- <span id="page-5-0"></span>1. Barbeira AN, Dickinson SP, Bonazzola R, Zheng J, Wheeler HE, Torres JM, et al. Exploring the phenotypic consequences of tissue specific gene expression variation inferred from GWAS summary statistics. Nat Commun. 2018;9(1):1825. DOI: [10.1038/s41467-018-03621-1.](https://doi.org/10.1038/s41467-018-03621-1) PMID: 29739930; PMCID: PMC5940825
- <span id="page-5-20"></span>2. Gusev A, Ko A, Shi H, Bhatia G, Chung W, Penninx BW, et al. Integrative approaches for large-scale transcriptome-wide association studies. Nat Genet. 2016;48(3):245–52. DOI: [10.1038/ng.3506.](https://doi.org/10.1038/ng.3506) PMID: 26854917; PMCID: [PMC4767558](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4767558)
- 3. Gamazon ER, Wheeler HE, Shah KP, Mozaffari SV, Aquino-Michaels K, Carroll RJ, et al. A gene-based association method for mapping traits using reference transcriptome data. Nat Genet. 2015;47(9):1091–8. DOI: [10.1038/ng.3367.](https://doi.org/10.1038/ng.3367) PMID: 26258848; PMCID: [PMC4552594](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4552594)
- <span id="page-5-1"></span>4. Hu Y, Li M, Lu Q, Weng H, Wang J, Zekavat SM, et al. A statistical framework for cross-tissue transcriptome-wide association analysis. Nat Genet.
- <span id="page-5-2"></span>5. Gandal MJ, Zhang P, Hadjimichael E, Walker RL, Chen C, Liu S, et al. Transcriptome-wide isoform-level dysregulation in ASD, schizophrenia, and [bipolar disorder. Science. 2018;362\(6420\):eaat8127. DOI:](https://doi.org/10.1126/science.aat8127) 10.1126/science. aat8127. PMID: 30545856; PMCID: [PMC6443102](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6443102)
- <span id="page-5-3"></span>6. Bhattacharya A, Vo DD, Jops C, Kim M, Wen C, Hervoso JL, et al. Isoform-level transcriptome-wide association uncovers genetic risk mechanisms for neuropsychiatric disorders in the human brain. Nature Genetics. 2023;55(12): 2117–28. DOI: [10.1038/s41588-023-01560-2.](https://doi.org/10.1038/s41588-023-01560-2) PMID: 38036788; PMCID: [PMC10703692](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10703692)
- <span id="page-5-4"></span>7. Cui Y, Peng F, Wang D, Li Y, Li JS, Li L, et al. 3'aQTL-atlas: an atlas of 3'UTR alternative polyadenylation quantitative trait loci across human normal tissues. Nucleic Acids Res. 2022;50(D1):D39–45. DOI: [10.1093/nar/gkab740.](https://doi.org/10.1093/nar/gkab740) PMID: 34432052; PMCID: [PMC8728222](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8728222)
- <span id="page-5-5"></span>8. Cui Y, Arnold FJ, Peng F, Wang D, Li JS, Michels S, et al. Alternative polyadenylation transcriptome-wide association study identifies APA-linked susceptibility [genes in brain disorders. Nat Commun. 2023;14\(1\):583. DOI:](https://doi.org/10.1038/s41467-023-36311-8) 10.1038/s41467- 023-36311-8. PMID: 36737438; PMCID: [PMC9898543](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9898543)
- <span id="page-5-6"></span>9. Rhinn H, Qiang L, Yamashita T, Rhee D, Zolin A, Vanti W, et al. Alternative α-synuclein transcript usage as a convergent mechanism in Parkinson's disease pathology. Nat Commun. 2012;3:1084. DOI: [10.1038/ncomms2032.](https://doi.org/10.1038/ncomms2032) PMID: 23011138; PMCID: [PMC3660047](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3660047)
- <span id="page-5-7"></span>10. Romo L, Ashar-Patel A, Pfister E, Aronin N. Alterations in mRNA 3' UTR isoform abundance accompany gene expression changes in human huntington's [disease brains. Cell Rep. 2017;20\(13\):3057–70. DOI:](https://doi.org/10.1016/j.celrep.2017.09.009) 10.1016/j.celrep.2017.09. 009. PMID: 28954224; PMCID: [PMC5625827](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5625827)
- <span id="page-5-8"></span>11. Gennarino VA, Alcott CE, Chen CA, Chaudhury A, Gillentine MA, Rosenfeld JA, et al. NUDT21-spanning CNVs lead to neuropsychiatric disease and altered MeCP2 abundance via alternative polyadenylation. Elife. 2015;4:e10782. DOI: [10.7554/eLife.10782.](https://doi.org/10.7554/eLife.10782) PMID: 26312503; PMCID: [PMC4586391](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4586391)
- <span id="page-5-9"></span>12. Derti A, Garrett-Engele P, MacIsaac KD, Stevens RC, Sriram S, Chen R, et al. A quantitative atlas of polyadenylation in five mammals. Genome Res. 2012;22(6):1173–83. DOI: [10.1101/gr.132563.111.](https://doi.org/10.1101/gr.132563.111) PMID: 22454233; PMCID: [PMC3371698](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3371698)
- <span id="page-5-10"></span>13. Garneau NL, Wilusz J, Wilusz CJ. The highways and byways of mRNA decay. Nat Rev Mol Cell Biol. 2007;8(2):113–26. DOI: [10.1038/nrm2104.](https://doi.org/10.1038/nrm2104) PMID: 17245413
- <span id="page-5-11"></span>14. Sandberg R, Neilson JR, Sarma A, Sharp PA, Burge CB. Proliferating cells express mRNAs with shortened 3'untranslated regions and fewer microRNA target sites. Science. 2008;320(5883):1643–7. DOI: [10.1126/science.1155390.](https://doi.org/10.1126/science.1155390) PMID: 18566288; PMCID: [PMC2587246](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2587246)
- <span id="page-5-12"></span>15. Bartel DP. MicroRNAs: target recognition and regulatory functions. Cell. 2009;136(2):215–33. DOI: [10.1016/j.cell.2009.01.002.](https://doi.org/10.1016/j.cell.2009.01.002) PMID: 19167326; PMCID: [PMC3794896](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3794896)
- <span id="page-5-13"></span>16. Ji Z, Lee JY, Pan Z, Jiang B, Tian B. Progressive lengthening of 3' untranslated regions of mRNAs by alternative polyadenylation during mouse embryonic de[velopment. Proc Natl Acad Sci U S A. 2009;106\(17\):7028–33. DOI: doi:10.1073/](https://doi.org/10.1073/pnas.0900028106) pnas.0900028106. PMID: 19372383; PMCID: [PMC2669788](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2669788)
- <span id="page-5-14"></span>17. Agarwal V, Lopez-Darwin S, Kelley DR, Shendure J. The landscape of alternative polyadenylation in single cells of the developing mouse embryo. Nat Commun. 2021;12(1):5101. DOI: [10.1038/s41467-021-25388-8.](https://doi.org/10.1038/s41467-021-25388-8) PMID: 34429411; PMCID: [PMC8385098](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8385098)
- <span id="page-5-15"></span>18. An JJ, Gharami K, Liao G-Y, Woo NH, Lau AG, Vanevski F, et al. Distinct role of long 3' UTR BDNF mRNA in spine morphology and synaptic plasticity in hippocampal neurons. Cell. 2008;134(1):175–87. PMID: 18614020; PMCID: [PMC2527207](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2527207) Consortium GT, Laboratory DA, Coordinating Center-Analysis Working G, Statistical Methods groups—Analysis Working G, Enhancing Gg, Fund NIHC, et al. Genetic effects on gene expression across human tissues. Nature. 2017;550(7675):204- 13. https://doi.org/10.1038/nature24277.
- <span id="page-5-16"></span>19. Consortium GT, Laboratory DA, Coordinating Center-Analysis Working G, Statistical Methods groups—Analysis Working G, Enhancing Gg, Fund NIHC, et al. Genetic effects on gene expression across human tissues. Nature. 2017;550(7675):204–13. DOI: [10.1038/nature24277.](https://doi.org/10.1038/nature24277) PMID: 29022597; PMCID: [PMC5776756](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5776756)
- <span id="page-5-17"></span>20. Bennett DA, Buchman AS, Boyle PA, Barnes LL, Wilson RS, Schneider JA. Religious orders study and rush memory and aging project. J Alzheimers Dis. 2018;64(s1):S161–89. DOI: [10.3233/JAD-179939.](https://doi.org/10.3233/JAD-179939) PMID: 29865057; PMCID: [PMC6380522](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6380522)
- <span id="page-5-18"></span>21. PsychENCODE Consortium, Akbarian S, Liu C, Knowles JA, Vaccarino FM, Farnham PJ, et al. The PsychENCODE project. Nat Neurosci. 2015;18(12):1707–12. DOI: [10.1038/nn.4156.](https://doi.org/10.1038/nn.4156) PMID: 26605881; PMCID: [PMC4675669](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4675669)
- <span id="page-5-19"></span>22. Xia Z, Donehower LA, Cooper TA, Neilson JR, Wheeler DA, Wagner EJ, et al. Dynamic analyses of alternative polyadenylation from RNA-seq reveal a 3'-UTR [landscape across seven tumour types. Nat Commun. 2014;5:5274. DOI:](https://doi.org/10.1038/ncomms6274) 10.1038/ ncomms6274. PMID: 25409906; PMCID: [PMC4467577](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4467577)
- <span id="page-6-0"></span>23. Gonzalez M, Nampoothiri S, Kornblum C, Oteyza AC, Walter J, Konidari I, et al. Mutations in phospholipase DDHD2 cause autosomal recessive hereditary spas[tic paraplegia \(SPG54\). Eur J Hum Genet. 2013;21\(11\):1214–8. DOI:](https://doi.org/10.1038/ejhg.2013.29) 10.1038/ ejhg.2013.29. PMID: 23486545; PMCID: [PMC3798837](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3798837)
- <span id="page-6-1"></span>24. Inloes JM, Hsu KL, Dix MM, Viader A, Masuda K, Takei T, et al. The hereditary spastic paraplegia-related enzyme DDHD2 is a principal brain triglyceride li[pase. Proc Natl Acad Sci U S A. 2014;111\(41\):14924–9. DOI:](https://doi.org/10.1073/pnas.1413706111) 10.1073/pnas. 1413706111. PMID: 25267624; PMCID: [PMC4205627](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4205627)
- <span id="page-6-2"></span>25. Zhang C, Li X, Zhao L, Guo W, Deng W, Wang Q, et al. Brain transcriptomewide association study implicates novel risk genes underlying schizophrenia risk. Psychol Med. 2023:1–11. DOI: [10.1017/S0033291723000417.](https://doi.org/10.1017/S0033291723000417) PMID: 37092861
- <span id="page-6-3"></span>26. Shi Y, Li Z, Xu Q, Wang T, Li T, Shen J, et al. Common variants on 8p12 and 1q24.2 [confer risk of schizophrenia. Nat Genet. 2011;43\(12\):1224–7. DOI:](https://doi.org/10.1038/ng.980) 10.1038/ng. 980. PMID: 22037555; PMCID: [PMC3773910](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3773910)
- <span id="page-6-4"></span>27. Pardiñas AF, Holmans P, Pocklington AJ, Escott-Price V, Ripke S, Carrera N, et al. Common schizophrenia alleles are enriched in mutation-intolerant genes and in regions under strong background selection. Nat Genet. 2018;50(3):381–9. DOI: [10.1038/s41588-018-0059-2.](https://doi.org/10.1038/s41588-018-0059-2) PMID: 29483656; PMCID: [PMC5918692](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5918692)
- <span id="page-6-5"></span>28. Matoba N, Liang D, Sun H, Aygün N, McAfee JC, Davis JE, et al. Common genetic risk variants identified in the SPARK cohort support DDHD2 as a candidate risk [gene for autism. Transl Psychiatry. 2020;10\(1\):265. DOI:](https://doi.org/10.1038/s41398-020-00953-9) 10.1038/s41398-020- 00953-9. PMID: 32747698; PMCID: [PMC7400671](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7400671)
- <span id="page-6-6"></span>29. Hall LS, Pain O, O'Brien HE, Anney R, Walters JTR, Owen MJ, et al. Cis-effects on gene expression in the human prenatal brain associated with genetic risk for [neuropsychiatric disorders. Mol Psychiatry. 2021;26\(6\):2082–8. DOI:](https://doi.org/10.1038/s41380-020-0743-3) 10.1038/ s41380-020-0743-3. PMID: 32366953; PMCID: [PMC7611670](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7611670)
- <span id="page-6-7"></span>30. Trubetskoy V, Pardiñas AF, Qi T, Panagiotaropoulou G, Awasthi S, Bigdeli TB, et al. Mapping genomic loci implicates genes and synaptic biology in schizophrenia. Nature. 2022;604(7906):502–8. DOI: [10.1038/s41586-022-04434-5.](https://doi.org/10.1038/s41586-022-04434-5) PMID: 35396580; PMCID: [PMC9392466](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9392466)
- <span id="page-6-8"></span>31. Dang X, Liu J, Zhang Z, Luo X-J. Mendelian randomization study using dopaminergic neuron-specific eQTL identifies novel risk genes for schizophrenia. Mol Neurobiol. 2023;60(3):1537–46. DOI: [10.1007/s12035-022-03160-3.](https://doi.org/10.1007/s12035-022-03160-3) PMID: 36517655
- <span id="page-6-9"></span>32. Park CY, Zhou J, Wong AK, Chen KM, Theesfeld CL, Darnell RB, et al. Genomewide landscape of RNA-binding protein target site dysregulation reveals a major impact on psychiatric disorder risk. Nat Genet. 2021;53(2):166–73. DOI: [10.1038/s41588-020-00761-3.](https://doi.org/10.1038/s41588-020-00761-3) PMID: 33462483; PMCID: [PMC7886016](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7886016)
- <span id="page-6-10"></span>33. Aberg K, Saetre P, Lindholm E, Ekholm B, Pettersson U, Adolfsson R, et al. Human QKI, a new candidate gene for schizophrenia involved in myelination. Am J [Med Genet B Neuropsychiatr Genet. 2006;141B\(1\):84–90. DOI:](https://doi.org/10.1002/ajmg.b.30243) 10.1002/ajmg.b. 30243. PMID: 16342280
- <span id="page-6-11"></span>34. Alkanderi S, Molinari E, Shaheen R, Elmaghloob Y, Stephen LA, Sammut V, et al. ARL3 mutations cause Joubert syndrome by disrupting Ciliary Protein composition. Am J Hum Genet. 2018;103(4):612–20. DOI: [10.1016/j.ajhg.2018.08.015.](https://doi.org/10.1016/j.ajhg.2018.08.015) PMID: 30269812; PMCID: [PMC6174286](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6174286)
- <span id="page-6-12"></span>35. Yu H, Yan H, Li J, Li Z, Zhang X, Ma Y, et al. Common variants on 2p16.1, 6p22.1 and 10q24.32 are associated with schizophrenia in Han Chinese population. Mol Psychiatry. 2017;22(7):954–60. DOI: [10.1038/mp.2016.212.](https://doi.org/10.1038/mp.2016.212) PMID: 27922604
- <span id="page-6-13"></span>36. Liu J, Li X, Luo XJ. Proteome-wide association study provides insights into the genetic component of protein abundance in psychiatric disorders. Biol Psychiatry. 2021;90(11):781–9. DOI: [10.1016/j.biopsych.2021.06.022.](https://doi.org/10.1016/j.biopsych.2021.06.022) PMID: 34454697
- <span id="page-6-14"></span>37. Ma L, Shcherbina A, Chetty S. Variations and expression features of CYP2D6 [contribute to schizophrenia risk. Mol Psychiatry. 2021;26\(6\):2605–15. DOI:](https://doi.org/10.1038/s41380-020-0675-y) 10. 1038/s41380-020-0675-y. PMID: 32047265; PMCID: [PMC8440189](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8440189)
- <span id="page-6-16"></span>38. Ma L, Semick SA, Chen Q, Li C, Tao R, Price AJ, et al. Schizophrenia risk variants influence multiple classes of transcripts of sorting nexin 19 (SNX19). Mol Psychiatry. 2020;25(4):831–43. DOI: [10.1038/s41380-018-0293-0.](https://doi.org/10.1038/s41380-018-0293-0) PMID: 30635639
- <span id="page-6-15"></span>39. Gusev A, Mancuso N, Won H, Kousi M, Finucane HK, Reshef Y, et al. Transcriptome-wide association study of schizophrenia and chromatin activ[ity yields mechanistic disease insights. Nat Genet. 2018;50\(4\):538–48. DOI:](https://doi.org/10.1038/s41588-018-0092-1) 10. 1038/s41588-018-0092-1. PMID: 29632383; PMCID: [PMC5942893](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5942893)
- <span id="page-6-17"></span>40. Takahashi Y, Maynard KR, Tippani M, Jaffe AE, Martinowich K, Kleinman JE, et al. Single molecule in situ hybridization reveals distinct localizations of schizophrenia risk-related transcripts SNX19 and AS3MT in human brain. Mol Psychiatry. 2021;26(7):3536–47. DOI: [10.1038/s41380-021-01046-9.](https://doi.org/10.1038/s41380-021-01046-9) PMID: 33649454
- <span id="page-6-18"></span>41. Nicolas E, Poitelon Y, Chouery E, Salem N, Levy N, Megarbane A, et al. CAMOS, a nonprogressive, autosomal recessive, congenital cerebellar ataxia, is caused by a mutant zinc-finger protein, ZNF592. Eur J Hum Genet. 2010;18(10):1107–13. DOI: [10.1038/ejhg.2010.82.](https://doi.org/10.1038/ejhg.2010.82) PMID: 20531441; PMCID: [PMC2987462](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2987462)
- <span id="page-6-19"></span>42. Ikeda M, Takahashi A, Kamatani Y, Okahisa Y, Kunugi H, Mori N, et al. A genome-wide association study identifies two novel susceptibility loci and trans population polygenicity associated with bipolar disorder. Mol Psychia-

try. 2018;23(3):639–47. DOI: [10.1038/mp.2016.259.](https://doi.org/10.1038/mp.2016.259) PMID: 28115744; PMCID: [PMC5822448](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5822448)

- <span id="page-6-20"></span>43. Zhao L, Chang H, Zhou DS, Cai J, Fan W, Tang W, et al. Replicated associations of FADS1, MAD1L1, and a rare variant at 10q26.13 with bipolar disorder in Chinese [population. Transl Psychiatry. 2018;8\(1\):270. DOI:](https://doi.org/10.1038/s41398-018-0337-x) 10.1038/s41398-018-0337 x. PMID: 30531795; PMCID: [PMC6286364](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6286364)
- <span id="page-6-21"></span>44. Stahl EA, Breen G, Forstner AJ, McQuillin A, Ripke S, Trubetskoy V, et al. Genome-wide association study identifies 30 loci associated with bipolar disorder. Nat Genet. 2019;51(5):793–803. DOI: [10.1038/s41588-019-0397-8.](https://doi.org/10.1038/s41588-019-0397-8) PMID: 31043756; PMCID: [PMC6956732](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6956732)
- <span id="page-6-22"></span>45. Mullins N, Forstner AJ, O'Connell KS, Coombes B, Coleman JRI, Qiao Z, et al. Genome-wide association study of more than 40,000 bipolar disorder cases provides new insights into the underlying biology. Nat Genet. 2021;53(6):817–29. DOI: [10.1038/s41588-021-00857-4.](https://doi.org/10.1038/s41588-021-00857-4) PMID: 34002096; PMCID: [PMC8192451](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8192451)
- <span id="page-6-23"></span>46. Park WJ, Kothapalli KS, Reardon HT, Lawrence P, Qian SB, Brenna JT. A novel FADS1 isoform potentiates FADS2-mediated production of eicosanoid precursor fatty acids. J Lipid Res. 2012;53(8):1502–12. DOI: [10.1194/jlr.M025312.](https://doi.org/10.1194/jlr.M025312) PMID: 22619218; PMCID: [PMC3540860](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3540860)
- <span id="page-6-24"></span>47. Ameur A, Enroth S, Johansson A, Zaboli G, Igl W, Johansson AC, et al. Genetic adaptation of fatty-acid metabolism: a human-specific haplotype increasing the biosynthesis of long-chain omega-3 and omega-6 fatty acids. Am J Hum Genet. 2012;90(5):809–20. DOI: [10.1016/j.ajhg.2012.03.014.](https://doi.org/10.1016/j.ajhg.2012.03.014) PMID: 22503634; PMCID: [PMC3376635](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3376635)
- <span id="page-6-25"></span>48. Yamamoto H, Lee-Okada HC, Ikeda M, Nakamura T, Saito T, Takata A, et al. GWASidentified bipolar disorder risk allele in the FADS1/2 gene region links mood episodes and unsaturated fatty acid metabolism in mutant mice. Mol Psychiatry. 2023;28(7):2848–56. DOI: [10.1038/s41380-023-01988-2.](https://doi.org/10.1038/s41380-023-01988-2) PMID: 36806390; PMCID: [PMC10615742](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC10615742)
- <span id="page-6-26"></span>49. Stoll AL, Severus WE, Freeman MP, Rueter S, Zboyan HA, Diamond E, et al. Omega 3 fatty acids in bipolar disorder: a preliminary double-blind, placebo-controlled trial. Arch Gen Psychiatry. 1999;56(5):407–12. DOI: [10.1001/archpsyc.56.5.407.](https://doi.org/10.1001/archpsyc.56.5.407) PMID: 10232294
- <span id="page-6-27"></span>50. Frangou S, Lewis M, McCrone P. Efficacy of ethyl-eicosapentaenoic acid in bipolar depression: randomised double-blind placebo-controlled study. Br J Psychiatry. 2006;188:46–50. DOI: [10.1192/bjp.188.1.46.](https://doi.org/10.1192/bjp.188.1.46) PMID: 16388069
- <span id="page-6-28"></span>51. Engin E, Liu J, Rudolph U.  $\alpha$ 2-containing GABA(A) receptors: a target for the development of novel treatment strategies for CNS disorders. Pharmacol Ther. 2012;136(2):142–52. DOI: [10.1016/j.pharmthera.2012.08.006.](https://doi.org/10.1016/j.pharmthera.2012.08.006) PMID: 22921455; PMCID: [PMC3478960](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3478960)
- <span id="page-6-29"></span>52. Edenberg HJ, Foroud T. Genetics and alcoholism. Nat Rev Gastroenterol Hepatol. 2013;10(8):487–94. DOI: [10.1038/nrgastro.2013.86.](https://doi.org/10.1038/nrgastro.2013.86) PMID: 23712313; PMCID: [PMC4056340](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4056340)
- <span id="page-6-30"></span>53. Enoch MA, Hodgkinson CA, Yuan Q, Albaugh B, Virkkunen M, Goldman D. GABRG1 and GABRA2 as independent predictors for alcoholism in two populations. Neuropsychopharmacology. 2009;34(5):1245–54. DOI: [10.1038/npp.2008.171.](https://doi.org/10.1038/npp.2008.171) PMID: 18818659; PMCID: [PMC2656604](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2656604)
- <span id="page-6-31"></span>54. Wellcome Trust Case Control Consortium. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature. 2007;447(7145):661–78. DOI: [10.1038/nature05911.](https://doi.org/10.1038/nature05911) PMID: 17554300; PMCID: [PMC2719288](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2719288)
- 55. Ren H, Guan L, Zhao L, Lin Y, Wang Y, Yang Z, et al. Contribution of genes in the GABAergic pathway to bipolar disorder and its executive function deficit in the Chinese Han population. Am J Med Genet B Neuropsychiatr Genet. 2018;177(1):50–67. DOI: [10.1002/ajmg.b.32601.](https://doi.org/10.1002/ajmg.b.32601) PMID: 29135068
- <span id="page-6-32"></span>56. Craddock N, Jones L, Jones IR, Kirov G, Green EK, Grozeva D, et al. Strong genetic evidence for a selective influence of GABAA receptors on a component of the [bipolar disorder phenotype. Mol Psychiatry. 2010;15\(2\):146–53. DOI:](https://doi.org/10.1038/mp.2008.66) 10.1038/ mp.2008.66. PMID: 19078961; PMCID: [PMC3967096](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3967096)
- <span id="page-6-33"></span>57. Barker JS, Hines RM. Regulation of GABA<sub>A</sub> receptor subunit expression in [substance use disorders. Int J Mol Sci. 2020;21\(12\):4445. DOI:](https://doi.org/10.3390/ijms21124445) 10.3390/ ijms21124445.
- <span id="page-6-34"></span>58. Schieweck R, Kiebler MA. Posttranscriptional gene regulation of the GABA receptor to control neuronal inhibition. Front Mol Neurosci. 2019;12:152. DOI: [10.3389/fnmol.2019.00152.](https://doi.org/10.3389/fnmol.2019.00152) PMID: 31316346; PMCID: [PMC6611381](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6611381)
- <span id="page-6-35"></span>59. Psychiatric GWAS Consortium Bipolar Disorder Working Group. Large-scale genome-wide association analysis of bipolar disorder identifies a new susceptibility locus near ODZ4. Nat Genet. 2011;43(10):977–83. DOI: [10.1038/ng.943.](https://doi.org/10.1038/ng.943) PMID: 21926972; PMCID: [PMC3637176](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3637176)
- <span id="page-6-36"></span>60. Ferreira MA, O'Donovan MC, Meng YA, Jones IR, Ruderfer DM, Jones L, et al. Collaborative genome-wide association analysis supports a role for ANK3 and [CACNA1C in bipolar disorder. Nat Genet. 2008;40\(9\):1056–8. DOI:](https://doi.org/10.1038/ng.209) 10.1038/ng. 209. PMID: 18711365; PMCID: [PMC2703780](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2703780)
- <span id="page-6-37"></span>61. Gray AC, Raingo J, Lipscombe D. Neuronal calcium channels: splicing for optimal [performance. Cell Calcium. 2007;42\(4-5\):409–17. DOI:](https://doi.org/10.1016/j.ceca.2007.04.003) 10.1016/j.ceca.2007.04. 003. PMID: 17512586; PMCID: [PMC2001240](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2001240)





- <span id="page-7-0"></span>62. Raingo J, Castiglioni AJ, Lipscombe D. Alternative splicing controls G proteindependent inhibition of N-type calcium channels in nociceptors. Nat Neurosci. 2007;10(3):285–92. DOI: [10.1038/nn1848.](https://doi.org/10.1038/nn1848) PMID: 17293861; PMCID: [PMC3027493](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3027493)
- <span id="page-7-1"></span>63. Marangoudakis S, Andrade A, Helton TD, Denome S, Castiglioni AJ, Lipscombe D. Differential ubiquitination and proteasome regulation of Ca(V)2.2 N-type chan[nel splice isoforms. J Neurosci. 2012;32\(30\):10365–9. DOI:](https://doi.org/10.1523/jneurosci.0851-11.2012) 10.1523/jneurosci. 0851-11.2012. PMID: 22836269; PMCID: [PMC3428229](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3428229)
- <span id="page-7-2"></span>64. Bunda A, LaCarubba B, Bertolino M, Akiki M, Bath K, Lopez-Soto J, et al. Cacna1b alternative splicing impacts excitatory neurotransmission and is linked to be[havioral responses to aversive stimuli. Mol Brain. 2019;12\(1\):81. DOI:](https://doi.org/10.1186/s13041-019-0500-1) 10.1186/ s13041-019-0500-1.
- <span id="page-7-3"></span>65. Schorge S, Gupta S, Lin Z, McEnery MW, Lipscombe D. Calcium channel activation stabilizes a neuronal calcium channel mRNA. Nat Neurosci. 1999;2(9):785–90. DOI: [10.1038/12153.](https://doi.org/10.1038/12153) PMID: 31630675; PMCID: [PMC6802325](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6802325)
- <span id="page-7-4"></span>66. Pazzaglia PJ, Post RM, Ketter TA, Callahan AM, Marangell LB, Frye MA, et al. Nimodipine monotherapy and carbamazepine augmentation in patients with refractory recurrent affective illness. J Clin Psychopharmacol. 1998;18(5)404–13. DOI: [10.1097/00004714-199810000-00009.](https://doi.org/10.1097/00004714-199810000-00009) PMID: 9790159
- <span id="page-7-5"></span>67. Li M, Yuan N, Nurnberger JI, Alliey-Rodriguez N, Zhou J, Duan F, et al. A pilot pharmacogenetic study of calcium channel blocker treatment of bipolar mania. Psychiatry Res. 2023;326:115281. DOI: [10.1016/j.psychres.2023.115281.](https://doi.org/10.1016/j.psychres.2023.115281) PMID: 37270865
- <span id="page-7-6"></span>68. Cheng W, van der Meer D, Parker N, Hindley G, O'Connell KS, Wang Y, et al. Shared genetic architecture between schizophrenia and subcortical brain volumes implicates early neurodevelopmental processes and brain development [in childhood. Mol Psychiatry. 2022;27\(12\):5167–76. DOI:](https://doi.org/10.1038/s41380-022-01751-z) 10.1038/s41380-022- 01751-z. PMID: 36100668
- <span id="page-7-7"></span>69. Mai H, Bao J, Thompson PM, Kim D, Shen L. Identifying genes associated with brain volumetric differences through tissue specific transcriptomic inference from GWAS summary data. BMC Bioinformatics. 2022;23(Suppl 3):398. DOI: [10.1186/s12859-022-04947-w.](https://doi.org/10.1186/s12859-022-04947-w) PMID: 36171548; PMCID: [PMC9520794](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9520794)
- <span id="page-7-8"></span>70. Zaltsman Y, Shachnai L, Yivgi-Ohana N, Schwarz M, Maryanovich M, Houtkooper RH, et al. MTCH2/MIMP is a major facilitator of tBID recruitment to mitochondria. Nat Cell Biol. 2010;12(6):553–62. DOI: [10.1038/ncb2057.](https://doi.org/10.1038/ncb2057) PMID: 20436477; PMCID: [PMC4070879](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4070879)
- <span id="page-7-9"></span>71. Buzaglo-Azriel L, Kuperman Y, Tsoory M, Zaltsman Y, Shachnai L, Zaidman SL, et al. Loss of muscle MTCH2 increases whole-body energy utilization and pro[tects from diet-induced obesity. Cell Rep. 2016;14\(7\):1602–10. DOI:](https://doi.org/10.1016/j.celrep.2016.01.046) 10.1016/j. celrep.2016.01.046. PMID: 26876167
- <span id="page-7-10"></span>72. Willer CJ, Speliotes EK, Loos RJ, Li S, Lindgren CM, Heid IM, et al. Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. Nat Genet. 2009;41(1):25–34. DOI: [10.1038/ng.287.](https://doi.org/10.1038/ng.287) PMID: 19079261; PMCID: [PMC2695662](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2695662)
- <span id="page-7-11"></span>73. Luciano M, Hagenaars SP, Davies G, Hill WD, Clarke TK, Shirali M, et al. Association analysis in over 329,000 individuals identifies 116 independent variants in[fluencing neuroticism. Nat Genet. 2018;50\(1\):6–11. DOI:](https://doi.org/10.1038/s41588-017-0013-8) 10.1038/s41588-017- 0013-8. PMID: 29255261; PMCID: [PMC5985926](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5985926)
- <span id="page-7-12"></span>74. Day FR, Ong KK, Perry JRB. Elucidating the genetic basis of social interaction and isolation. Nat Commun. 2018;9(1):2457. DOI: [10.1038/s41467-018-04930-1.](https://doi.org/10.1038/s41467-018-04930-1) PMID: 29970889; PMCID: [PMC6030100](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6030100)
- <span id="page-7-13"></span>75. Manjunath LE, Singh A, Sahoo S, Mishra A, Padmarajan J, Basavaraju CG, et al. Stop codon read-through of mammalian *MTCH2* leading to an unstable isoform regulates mitochondrial membrane potential. J Biol Chem.

2020;295(50):17009–26. DOI: [10.1074/jbc.RA120.014253.](https://doi.org/10.1074/jbc.RA120.014253) PMID: 33028634; PMCID: [PMC7863902](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7863902)

- <span id="page-7-14"></span>76. Jensen LJ, Kuhn M, Stark M, Chaffron S, Creevey C, Muller J, et al. STRING 8– a global view on proteins and their functional interactions in 630 organisms. Nucleic Acids Res. 2009;37(Database issue):D412–6. DOI: [10.1093/nar/gkn760.](https://doi.org/10.1093/nar/gkn760) PMID: 18940858; PMCID: [PMC2686466](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2686466)
- <span id="page-7-15"></span>77. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res. 2003;13(11):2498–504. DOI: [10.1101/gr.1239303.](https://doi.org/10.1101/gr.1239303) PMID: 14597658; PMCID: [PMC403769](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC403769)
- <span id="page-7-16"></span>78. Altaf-Ul-Amin M, Hirose K, Nani JV, Porta LC, Tasic L, Hossain SF, et al. A system biology approach based on metabolic biomarkers and protein–protein interactions for identifying pathways underlying schizophrenia and bipolar disorder. Sci Rep. 2021;11(1):14450. DOI: [10.1038/s41598-021-93653-3.](https://doi.org/10.1038/s41598-021-93653-3) PMID: 34262063; PMCID: [PMC8280132](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8280132)
- <span id="page-7-17"></span>79. Amir RE, Van den Veyver IB, Wan M, Tran CQ, Francke U, Zoghbi HY. Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2. Nat Genet. 1999;23(2):185–8. DOI: [10.1038/13810.](https://doi.org/10.1038/13810) PMID: 10508514
- <span id="page-7-18"></span>80. Coy JF, Sedlacek Z, Bachner D, Delius H, Poustka A. A complex pattern of evolutionary conservation and alternative polyadenylation within the long 3" untranslated region of the methyl-CpG-binding protein 2 gene (MeCP2) suggests a regulatory role in gene expression. Hum Mol Genet. 1999;8(7):1253–62. DOI: [10.1093/hmg/8.7.1253.](https://doi.org/10.1093/hmg/8.7.1253) PMID: 10369871
- <span id="page-7-19"></span>81. Balmer D, Goldstine J, Rao YM, LaSalle JM. Elevated methyl-CpG-binding protein 2 expression is acquired during postnatal human brain development and is correlated with alternative polyadenylation. J Mol Med (Berl). 2003;81(1):61–8. DOI: [10.1007/s00109-002-0396-5.](https://doi.org/10.1007/s00109-002-0396-5) PMID: 12545250
- <span id="page-7-20"></span>82. Millevoi S, Vagner S. Molecular mechanisms of eukaryotic pre-mRNA 3' end pro[cessing regulation. Nucleic Acids Res. 2010;38\(9\):2757–74. DOI:](https://doi.org/10.1093/nar/gkp1176) 10.1093/nar/ gkp1176. PMID: 20044349; PMCID: [PMC2874999](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2874999)

**Publisher's note:** Genomic Press maintains a position of impartiality and neutrality regarding territorial assertions represented in published materials and affiliations of institutional nature. As such, we will use the affiliations provided by the authors, without editing them. Such use simply reflects what the authors submitted to us and it does not indicate that Genomic Press supports any type of territorial assertions.

**Open Access.** This article is licensed to Genomic Press under the Cre-<u>@0®0 </u> ative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License (CC BY-NC-ND 4.0). The license mandates: (1) Attribution: Credit must be given to the original work, with a link to the license and notification of any changes. The acknowledgment should not imply licensor endorsement. (2) NonCommercial: The material cannot be used for commercial purposes. (3) NoDerivatives: Modified versions of the work cannot be distributed. (4) No additional legal or technological restrictions may be applied beyond those stipulated in the license. Public domain materials or those covered by statutory exceptions are exempt from these terms. This license does not cover all potential rights, such as publicity or privacy rights, which may restrict material use. Third-party content in this article falls under the article's Creative Commons license unless otherwise stated. If use exceeds the license scope or statutory regulation, permission must be obtained from the copyright holder. For complete license details, visit https://creativecommons.org/ [licenses/by-nc-nd/4.0/. The license is provided without warranties.](https://creativecommons.org/licenses/by-nc-nd/4.0/)